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(54) Title: PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

#### (57) Abstract

By this invention, compositions and methods of use related to  $\beta$ -ketoacyl-ACP synthase of special interest are synthases obtainable from *Cuphea* species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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WO 98/46776 PCT/US98/07114

# PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

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#### INTRODUCTION

#### Field of Invention

The present invention is directed to genes encoding plant fatty acid synthase enzymes relevant to fatty acid synthesis in plants, and to methods of using such genes in combination with genes encoding plant medium-chain preferring thioesterase proteins. Such uses provide a method to increase the levels of medium-chain fatty acids that may be produced in seed oils of transgenic plants.

#### Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

10 Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large

25 amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pqs. 111-133).

#### DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea

hookeriana KAS factor B clone chKAS B-2 are provided.

Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided.

Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided.

Figure 4. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-1-6 are provided.

Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided.

Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided.

Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided.

Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

- Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.
- Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- 5 Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
  - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- Figure 13. The activity profile for purified castor KAS factor B using various acyl-ACP substrates is provided. Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS

15 A-2-7 is provided.

- Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- 20 Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 17. Graphs showing the %C10/%C8 ratios in transgenic 25 plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.
  - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

WO 98/46776 PCT/US98/07114

plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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#### SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from Cuphea species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. KAS I class is sensitive to inhibition by cerulenin at concentrations as low as 1 µM. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50µM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

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Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell,

20 especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an anti
25 sense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

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#### DETAILED DESCRIPTION OF THE INVENTION

A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

PCT/US98/07114

activity towards acyl-ACPs having longer carbon chains,  $C_{14}$ - $C_{16}$ , and is inhibited by concentrations of cerulenin (50 $\mu$ M). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains,  $C_2$  to  $C_6$ , and is insensitive to inhibition by cerulenin.

Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles between the factor B synthase proteins from Cuphea and castor. The synthase A clone from castor, however, demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in

transgenic plant seeds which normally do not produce mediumchain fatty acids does not result in any detectable
modification of the fatty acid types and contents produced
in such seeds. However, when Cuphea hookeriana KAS A
protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, coexpression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, 25 an increased proportion of C12 fatty acids may be obtained by co-expression of Uc FatBl thioesterase and a chKAS A synthase factor proteins.

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However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatA1, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatA1 and plants expressing the Cuphea hookeriana KAS A protein.

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Thus, the instant invention provides methods of 15 increasing and/or altering the medium-chain fatty acid compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved 20 depending upon the particular fatty acids desired. For example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, 25 thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

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Recombinant constructs containing a nucleic acid sequence encoding a synthase protein factor or nucleic acid sequences encoding a synthase protein factor and a medium-chain acyl-ACP thioesterase may be prepared by methods well known in the art. Constructs may be designed to produce synthase in either prokaryotic or eukaryotic cells. The increased expression of a synthase in a plant cell, particularly in conjunction with expression of medium-chain thioesterases, or decreasing the amount of endogenous synthase observed in plant cells are of special interest.

Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the

transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes".

Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli,

B. subtilis, Saccharomyces cerevisiae, including genes such as ß-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of

transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of 20 the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

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Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

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For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

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#### EXAMPLES

#### Example 1 Cuphea KAS Factor A and B Gene Cloning

Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. For cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide The first 39 amino acids of cpuKAS B/8-7A are believed to represent the transit peptide, with the mature protein 20 encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A is provided. The mature protein is believed to begin at the lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

Deduced amino acid sequence of the C. hookeriana KAS 10 factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and 15 Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor 20 B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. The C. pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

#### Example 2 Levels and Patterns of Expression

mRNA and its abundance on the blot.

To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues 15 examined, whereas KAS A expression is detected only in the seed. These results also demonstrate a different level of expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65\_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and The larger hybridizing band is likely the transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA 25 screening corresponds well with the apparent mobility of the

## Example 3 Expression of Plant KAS Genes in E.coli

DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima

5 KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in

10 ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. The data demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

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The activity profile of the *C. hookeriana* KAS A clones

25 chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The *C. hookeriana* KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. In comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. The preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

#### Example 4 KAS and TE Expression in Transgenic Seed

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Both the CpFatB1 (*C. hookeriana* thioesterase cDNA;

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (*Pl.Mol.Biol.* (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

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A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (C. hookeriana thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-10 172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

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Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence 10 of two separate populations of heterozygotes. containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is 20 present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

hemizygous line led to an accumulation of up to 57 mol%
C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels
obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatA1, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% Cl8:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of Cl8:0 were reduced to approximately 12 mol%. Furthermore, levels of Cl6:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

#### Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µl) contained 0.1M Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10  $\mu M$  [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic *Brasica* (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

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extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS

A-2-7 also is a cerulenin-resistant condensing enzyme.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains.

- 15 All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.
- Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

factor protein heterologous to said transgenic plant in conjunction with expression of said plant medium-chain thioesterase, whereby the percentage of medium-chain fatty acids produced in seeds expressing both a plant synthase factor protein and a plant medium-chain thioesterase protein is increased as compared to the percentage of medium-chain fatty acids produced in seeds expressing only said plant medium-chain

- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatB1 protein.

thioesterase protein.

- 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
  - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.

- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a Cl2 preferring thioesterase from California bay.
- 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 25 1.
  - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
  - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

- 29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.
- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty 10 acid is C12 and said decreased fatty acid is C14.

48	96	144	192	240	288	336	384
GGC G1y	AAG Lys	GGT Gly	CAC	666 61y	TCA Ser	GCT Ala	ACT Thr
CCG Pro	TCC	GGT Gly	GGT Gly	ATG Met	TAT Tyr	GCC Ala	66C 61y
CCC	CTC	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala	GGA G1y
GAT Asp	CGC	GGA Gly	GAG Glu	ACA Thr	CCA Pro	CAT	GCT Ala
GTG Val	GAC Asp	ACA Thr	ATC Ile	ATT Ile	GGC G1у	TTC	ATT Ile
CTA Leu	GCC Ala	GGA Gly	CTT Leu	GCC Ala	ATG Met	TGC Cys	ATG Met
GAA Glu	GGT	GTC Val	TCT Ser	TAT Tyr	CTC	TAC	CTT
CTA Leu	CTC	CTG	CAG Gln	CCC Pro	GGT Gly	AAC Asn	GAT Asp
GCT Ala	GAT Asp	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC Ser	GCT Ala
GCC Ala	GCC	GGA Gly	666 61y	TTC	GAA Glu	ACT Thr	GAG Glu
GCG Ala	CGA Arg	GCC Ala	GAC Asp	TTC	ATC Ile	GCC	${ t GGT}$
GTG Val	GCA	AGA Arg	TCT Ser	CCT	GCT Ala	TGT Cys	CGT Arg
GCG Ala	TCG Ser	GAG Glu	TTC	ACC	CTC	GCA Ala	CGC Arg
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC Ile	CTG	ACT	ATC Ile
TCC	AGG Arg	GAC Asp	ACT	AAA Lys	GCC Ala	TCC Ser	CAT His
AGC	TGC Cys	ATC Ile	CTG	CGG	TCT	ATT Ile	AAT Asn

FIGURE 1 1 OF 4

432	480	528	576	624	672	720	768
AGG Arg	TGG	TTG Leu	ATT Ile	ACT Thr	AGC Ser	GCT Ala	ATC Ile
TGC	CCC	GTG Val	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC
GCT Ala	AGG Arg	GGA Gly	CCG	CAC His	GAG Glu	ATA Ile	AAT
GTG Val	TCT Ser	GCT Ala	GCA Ala	тат Ту <i>к</i>	ATT Ile	TAC	ATA Ile
TTT Phe	GCC Ala	GGT Gly	GGA Gly	GCT Ala	TGC Cys	AAT	GAG
66C 61y	ACT	GAA Glu	CGA Arg	GAT	TCT Ser	GTC	GCC
GGA G1y	CAG Gln	GGT Gly	AGA Arg	TGT Cys	TCT Ser	GAG Glu	
TTG Leu	CCG	ATG Met	ATG Met	AAC Asn	GTC Val	GAA Glu	GAT CTC Asp Leu FIGURE 1
GGG G1γ	GAC Asp	GTG Val	GCA Ala	ATC Ile	GGT Gly	CCT	666 61y
ATT Ile	GAT Asp	TTT Phe	CAT His	GCA Ala	CTT	TCA	GCT Ala
CCA	AAC Asn	$_{\rm G1Y}^{\rm GGT}$	GAA Glu	$_{\rm G1Y}$	GGT Gly	GTC Val	CTA
ATT Ile	AGG Arg	GAT Asp	TTG Leu	GGA G1y	GAT	66c 61y	ACT Thr
ATC Ile	CAA Gln	CGT Arg	AGC Ser	TTG Leu	GCT Ala	GCT Ala	TCT
GCA Ala	TCT	GAC Asp	GAG Glu	TAT Tyr	AGG Arg	GAT Asp	ACT
GCC	TTG	AAA Lys	ATG Met	GAG Glu	CCA	GAA Glu	GCG Ala
GAG Glu	GCT	GAT	GTG Val	GCA	GAT Asp	CTT	CAT

FIGURE 1 3 OF 4

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1236	1296	1348
TCTATGTAAT AAAACTAAGG ATTATTAATT TCCCTTTTAA TCCTGTCTCC AGTTTGAGCA	TGAAATTATA TTTATTTTAT CTTAGAAAGG TCAAATAAGA TTTTGTTTTA CCTCTGTAAA 1296	AA
TCCTGTCTCC	TTTTGTTTTA	ACTITITGITI GIAITGGAAA GGAAGIGCCG ICICAAAAAA AAAAAAAAA AA
TCCCTTTTAA	TCAAATAAGA	TCTCAAAAAA
ATTATTAATT	CTTAGAAAGG	GGAAGTGCCG
AAAACTAAGG	TTTATTTTAT	GTATTGGAAA
TCTATGTAAT	TGAAATTATA	ACTTTTGTTT

Sequence Range: 1 to 1704

_											
40 GTG Val>		GCA Ala>		TCT Ser>	190	GAC Asp>	240	cGG Arg>	CTC Leu>		GAA Glu>
GNG		TCG	140	GAC	1.5	ATC Ile		ATC Ile	AGG Arg		
ACC	90	AAT Asn	•	GTC Val		TTA		CAG Gln	30 AGG Arg	330	GCT Ala
30 TCC Ser		AGG Arg		GAC Asp		AGC	230	GGC Gly	280 GAC AGG ASP Arg		AAG Lys
AGC		TGC	130	TCC	180	ATC Ile	(1	GGC Gly	AAC Asn		AAG Lys
TGG	80	$_{\rm GGC}$	H	66C 61y		$_{\rm GGG}$		TTC	AAG Lys	320	GCC GGG A
20 AGC Ser		CCG Pro		TTC Phe		GAG AGC Glu Ser	220	AGG Arg	270 GGG Gly	(M)	GCC
AAA Lys		CCC		GTA Val	170	GAG Glu	22	ACC Thr	GAC		GTC Val
AAC	7.0	GAT Asp	120	TCC	**	GGC G1y		CCC	ATC Ile	0.	TGC ATT Cys Ile
10 AAA GGG Lys Gly	•	GTG Val		GTC		TCC		TTC	260 TAC Tyr	310	TGC
		CTA Leu		CTC	160	CTC	210	AAG Lys	GGA Gly		TAC
ACT		GAA Glu	110	GGC Gly	7(	CTC		TCC	ACG Thr		CGC Arg
CTC	<b>6</b> 09	CTA Leu		ATG GGC (Met Gly )		AAG Lys		GCT Ala	50 GCG Ala	300	CTC
ACC		GCT Ala		GGC Gly		GAA Glu	200	GAC Asp	250 AAC GCG Asn Ala		TGC
TTA Leu		GCC Ala	100	GCC Ala	150	TAC	(4	TTC Phe	TTC Phe		GAT Asp
AAA Lys	20	GCG	1(	CGA Arg		TAT Tyr		CGC	GGA Gly	90	GAC

FIGURE 2 1/5

											-	
	AGA Arg>	3.0	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	0	ATT Ile>
380	GAG Glu	43	${ m TTC}$		TCC	CTT Leu		GCA Ala	620	CGC Arg	670	ATC Ile
	AAG Lys		GTC Val		ATC Ile	520 GCT CTG Ala Leu	570	ACT Thr	Ū	ATC Ile	•.	GCA Ala
	GAT Asp		ACC Thr	470	AAG Lys			TCA		CAT His		GCT
370	ATT Ile	420	CTA	,	CGG Arg	TCT		ATT Ile	610	AAT Asn	<b>660</b>	GAG Glu
m	AAG Lys		66C 61y		CAC His	666 61y	260	TCG	6.3	GCC		ACT
	TCC		$_{\rm G1y}^{\rm GGT}$	460	AAA GGT Lys Gly	510 ATG Met	Δ,	TAT Tyr		GCT Ala		GGA Gly
	CTC	410	ATG Met	4		AAC		AAC		GCC Ala	650	GGA Gly
360	AGC		$_{\rm G1y}^{\rm GGT}$		GAG Glu	ACA Thr	550	CCA Pro	009	TAT Tyr	•	GCT
	GAA Glu		ACT Thr		ATC Ile	500 ATT Ile	5.	GGC Gly		TTT Phe		ATT Ile
	GGT Gly	400	GGA Gly	450	CTC	GCC		ATG Met		TGC	640	ATG Met
350	GGC Gly	4	GTT Val		AAT Asn	TAT Tyr		CTG	290	$\mathtt{TAC}$	64	CTC
	CTC		CTA Leu		CAG Gln	30 CCC Pro	540	$_{\rm GGT}$	• •	AAC Asn		GAC
	GAT Asp		GTG	440	GTT Val	490 ATT CC		TTG		TCC		GCT
340	TCC Ser	390	GGA G1y	•	666 61y	TTC Phe		GAT Asp	0	ACT Thr	630	GAG Glu
m	AAT Asn		GCT Ala		GAC ASP	TTT Phe	30	ATC Ile	580	GCT		GGC Gly

FIGURE 2 2/5

720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>	0.	GAT Asp>	096	GGG G1y>	ACT Thr>
	CAA Gln	CGT Arg		AGC	860	TTG	910	GCT		GCT	TCC
	TCT Ser	760 AAG GAC Lys Asp	810	GAG Glu	w	TAT		AGG Arg		GAT Asp	OO ACT Thr
710	TTA	76 AAG Lys		ATG Met		GAA Glu		CCA Pro	950	GAA Glu	1000 GCG AC Ala Th
• .	GCT Ala	GAT Asp		GTT Val	850	GCA	006	GAT Asp	01	CTG	CAT His
	AGG Arg	TGG Trd	800	TTG	8	ATT		ACT Thr		AGT	GCT
700	TGC	750 CCG Pro	w	GTA Val		ATT Ile		ATG Met	940	AGC	990 AAT Asn
7(	GCC Ala	AGG Arg		GGA Gly		CCG	890	CAT His	6	GAG Glu	ATA Ile
	GTT Val	TCA Ser	190	GCT	840	GCG Ala	w	TAT Tyr		ATT Ile	TAC
	TTC	740 GCC	7.5	$\begin{array}{c} GGG\\ G1Y \end{array}$		GGA Gly		GCT Ala		TGC	980 AAT Asn
069	GGA Gly	ACI		GAA Glu		CGA Arg	880	GAT Asp	930	TCT Ser	980 GTC AAT Val Asn
	GGA Gly	CAG		${\tt GGC}$	830	AAA Lys	88	TGT Cys		TCC	GAG Glu
	TTA	730 GAC CCT ASP Pro	780	ATG Met	w	ATG Met		AAT Asn		GTC Val	o GAA Glu
089	666 Gly			GTG Val		GCA Ala		GTC Val	920	$_{\rm GGT}$	970 CCT GAA Pro Glu
	ATT Ile	GAT Asp		TTT Phe	820	CAT His	870	GCA	01	CTT Leu	TCA
	CCA	AAT Asn	70	$_{\rm G1y}^{\rm GGT}$	8	GAA Glu		GGT Gly		$^{\rm GGG}_{\rm G1y}$	GTC Val

FIGURE 2 3/5

	AAG Lys>		CAC His>	0.9	GGA Gly>	1200	GAG Glu>	GAA Glu>		TCA Ser>		GCA
	TTC	1100	GGA Gly	1150	AAG Lys	•	CCC	CAT His		AAC Asn	1340	AAT
1050	GTT Val	ä	ATC Ile		ATT Ile		AAT Asn	10 CAA Gln	1290	CAC AAC	13	TCA AAT
•••	AAG GTT : Lys Val 1		ATG Met		ACA Thr	1190	CAA TTC Gln Phe	1240 CAG C2 Gln G3	-	GGC Gly		GGT
	AAG Lys	06	TCG	1140	GCG Ala	7		AAG Lys		GGA Gly	0 5	CTC
1040	ATC Ile	1090	AAG Lys		ATT Ile		AAC	AAG Lys	1280	TTC	1330	TTA
1(	GCC Ala		ACT		GCC Ala	30	AGC ATA Ser Ile	1230 AAC Asn	12	GGA Gly		TGA
	GAG ATA AAT Glu Ile Asn		AAT GCA Asn Ala	1130	GAA Glu	1180	AGC	GCC Ala		TTC		CCA Pro
30	ATA Ile	1080	AAT	H	CTT		CCC	GTT Val	0,	AAT TCA Asn Ser	1320	TTC AAG Phe Lys
1030	GAG Glu	• •	ATC Ile		GGT Gly		CAT His	1220 GAC ACA ASP Thr	1270			TTC
	GCC Ala		ACA Thr	30	$^{\rm GGG}_{\rm G1y}$	1170	CTT	12 GAC Asp		TCA		GCC
	CTT	1070	ATC Ile	1120	TCA		TGG Trp	TTC		ATC Ile	1310	TTC TCA Phe Ser
1020	GAT Asp	7(	GAA Glu		GCA Ala		GGC Gly	1210 GTG GAA Val Glu	1260	GCT Ala	13	TTC. Phe
•	$_{\rm GGG}$		AAG Lys		GGA Gly	1160	ACC Thr	1210 GTG GZ Val G	П	GTT Val		GCT Ala
	GCT Ala	20	ACC Thr	1110	CTT	ਜ	ACC Thr	TCA		AAT Asn	00	GTA Val
10	CTT	1060	AAC	• •	${\rm TGT} \\ {\rm Cys}$		ATA Ile	CCA '	20	GTG Val	1300	GTT Val

FIGURE 2 4/5

FIGURE 2 5/5

AATTTGTTGC TGAGACAGTG AGCTTCAACT TGCAGAGCAA TTTTTTACAT GCCTTGTCGT CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG TATTAGAAAG AACGAGGAA GATTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAAA AAAACTCGAG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG

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09	GCGGTGGCGG CCGCTCTAGA ACTAGTGGAT	120	GCTCAGGTGT	TGT ACG TGG Cys Thr Trp		CGT TCC Arg Ser	260	CTC TCC	310	CCT TGC Pro Cys	360	TTC GGA Phe Gly	
20	raga	110	GGTCGGCTCA	160 TTC TG Phe Cy	210	CCA Pro	(1)	ACT Thr		GAT Asp		CTC	٠.
	CTC		rcgg	16 CCT TT Pro Ph		GAC ASp		3 AGG J Arg		CTC	350	TCC	
	Ö			n O H		GAC AAC Asp Asn		CGG Arg	300	TGC	(*)	GCT	
40	9909	100	CTTG	G TCC a Ser	200	GAC	250	CGC		CAA Gln		TTC Phe	
	GGTG		TTCTTACTTG	150 T GCG 1 Ala	2	TCC		CGT Arg		TTC		GGA Gly	
0		0		GTT Val		TCA		TCC Ser	0	ACC	340	AAC Asn	
30	GAGCTCCACC	90	GGCACGAGTT	140 GCT TCT TGC ATG Ala Ser Cys Met		ACT Thr	240	CTC	290	TCC		GAT Asp	
	AGCT		GCAC	140 TTGC rCys	190	CCC	•••	CGC Arg		GGA Gly		GGG Gly	
20		80		14 F TCT a Ser	٠	ATG Met		CTC		CGC Arg	330	CTC	
	AAGC		GAAT			TGC	230	CGG Arg	280	CTC	(-)	TTC Phe	
	ACTAAAGGGA ACAAAAGCTG		GCAGGAATTC	0 G ACC a Thr	180	GCA		AAG Lys		TCC		CGC Arg	
10	GGA	70	GCT	130 G GCG t Ala		GCT		CAC His		TGC	320	CAA Gln	
	AAAG		CCCCGGGCT	A ATG Met		GTA Val		TCC	270	CAT His	33	CAG Gln	
	ACT		သသ	TCCA	170	CTC	220	CTT Leu	• •	TCC		AAC Asn	

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ACT Thr		GAA Glu		GTG Val		TAC	009	AAC Asn	TCT		GAC Asp
CGC		CAG Gln	200	GTT Val	550	GTT Val	9	GAG Glu	AAG Lys		ATG
66C 61y	450	GCA Ala	2(	GTA Val		GAT Asp		ATA Ile	ATC Ile	069	AGG Arg
400 CTC Leu	•	CCT		CGA Arg		CCC Pro	290	GAG Glu	640 GAG Glu	Φ	GAG Glu
AGG		CAA Gln		AGG Arg	540	gac Asp	50	AGT Ser	GGA Gly		TCC
CTG	440	ATG Met	490	CAA	υ,	CAT His		ATA Ile	GCC Ala	089	TTC
390 CAC His	4	GCT		AAG Lys		GGC Gly		AGT GGC Ser Gly	630 ATT Ile	99	AAG Lys
GGC G1y		GTG Val		ACC	530	CTA Leu	580	AGT	6 AGA Arg		CCA
CGC		ATG GCT Met Ala	480	GCT	2)	CCT		ATA Ile	ACG Thr		GCC
380 TCA AAT Ser Asn	430	ATG Met	•	CCT Pro		ACT Thr		GAC GGA ASP Gly	620 T CCC e Pro	670	GTG
		GTC Val		AAA Lys		GTG Val	570	GAC	62 TTT Phe		TGG Trp
CGT		GAG Glu	470	AAG Lys	520	GTG Val	u,	CTA	CAG Gln		66C 61y
CTT Leu	420	GGG Gly	4.	AAT Asn		GGC Gly		CTC	TCT Ser	099	GAT Asp
370 CCT Pro	•	TCC	,	ACA Thr		ATG Met	260	AAT Asn	610 TGC Cys	•	ACA
AAG Lys		CAT His		TCC Ser	510	$_{\rm G1y}^{\rm GGT}$	56	AAC Asn	GAC		TCC Ser
TCC	410	TCC	460	GTC Val	u,	ACA Thr		TAC	TTC Phe	650	TTT Phe

0	GAT Asp		TGT Cys	840	GAT Asp	TGT Cys		GAC		ACA Thr		GAA Glu
740	GCA Ala	790	AAG Lys	w	AGC	TTT Phe		ATG Met	980	GCA Ala	1030	GGC Gly
	TTA		AGA Arg		TTC	CCC	930	GCA	96	TGT Cys	<u> </u>	AAA Lys
	GCA Ala		AAA Lys	830	GTA Val	880 AGT Ser	01	CTT		GCC Ala		ATC Ile
730	AAA Lys	780	AAT Asn	8	AAG Lys	ATC Ile		ATT Ile		ACT Thr	1020	ATA Ile
•	AAG Lys		CTC		ATG	AAG Lys	920	GCT	970	TCA	10	CAC His
	66C Gly		GAG		GGT Gly	870 AAG Lys	. 6	TCC		ATA Ile		AAC Asn
720	GCA	770	AAA Lys	820	66C 61y	TAT Tyr		GGA Gly		TCG	9	GCG Ala
7	ACT	7.	ATG Met		TTG Leu	TCA		ATG Met	096	TAT Tyr	1010	GCT Ala
	CTG		GCG		GGA Gly	860 AGG ACT Arg Thr	910	AAT Asn		AAC Asn		AAT Asn
0	* TAC ATG Tyr Met		GAT Asp	810	TCC			ACA Thr		CCT		CTG
710		760	GAA Glu	~	66C 61y	CTG		ACC	950	ATG GGC Met Gly	1000	ATA Ile
	CTT Leu		ACT Thr		ATT Ile	GCT Ala	006	TCT	99		•	TGT
	ATG		ATC Ile	800	CTC	850 GAA Glu	01	TTT Phe		TGG Trp		TTC Phe
700	TTC Phe	750	GGA Gly	8	GTT Val	ATT Ile		CCT Pro		GGA Gly	066	AAC Asn
•	AAG Lys	•	GGT Gly		GGA Gly	TCC	890	GTA Val	940	TTG	o	AGT Ser

FIGURE 3 3 OF 6

1080	GTT Val	AAT Asn		TTT Phe		CAT His		AGT	1320	GCT Ala	TCG
Ħ	CCT	AAT Asn		GGA Gly	0	GAG Glu	1270	$\frac{\text{GGG}}{\text{Gly}}$	13	GGA Gly	GTC Val
	TTA Leu	AGG Arg	1170	GAT Asp	1220	TTA Leu	<b>~</b>	GGT		GAA Glu	GGA Gly
0.2	GTT Val	1120 CAG Gln	H	CGT Arg		GAG Glu		CTA Leu	0	CCT Pro	1360 TCC
1070	GCC	TCA		AAT Asn		GAG Glu	1260	TTT Phe	1310	CAC	cAG Gln
	GCG	TTG	20	AGT	1210	CTT	13	GAA Glu		CCT	GCT
	GAT	1110 CGA GCT Arg Ala	1160	GAC	• •	CTT		GCG		GAG Glu	1350 C TTG a Leu
1060	TCG			TGG Trp		TTA Leu	20	TAT TYE	1300	ATG ACC Met Thr	A)
	GGC Gly	TGC		CCA Pro	1200	GGA GTT Gly Val	1250	ATT ' Ile '	•	ATG Met	AAG Lys
	$_{\rm GGT}^{\rm GGT}$	1100 TA GCA	1150	AGA Arg	1;			ACC		CAC	1340 ATA GAG Ile Glu
1050	TGT Cys	0 >		TCG		GCT Ala		GCA Ala	1290	TAC	1340 ATA G Ile G
1	CTT	TTC		GCT	06	$_{\rm GGA}$	1240	GGT Gly	ä	GCC	TGC Cys
	ATG Met	.090 GGA GGT Gly Gly	1140	AAA Lys	1190	GAA Glu	` '	AGA Arg		GAC Asp	CTC
1040	* GAC ATG ASP Met		Н	ACC Thr		GGA Gly		AAA Lys	30	TGC	1330 ATC Ile
10		TTG		CCT		ATG Met	1230	AAG Lys	1280	ACT Thr	1 GTG Val
	GCA	GGT Gly	1130	GAC Asp	1180	GTG Val	ï	GCA		TTC	GGT Gly

FIGURE 3 4 OF 6

AGG GAA GAC GTA AAT TAC ATA AAT GCG CAT GCA ACT TCC ACT CCT GCT Arg Glu Asp Val Asn Tyr Ile Asn Ala His Ala Thr Ser Thr Pro Ala

GGA GAT ATC AAG GAA TAC CAA GCT CTC GCC CAC TGT TTC GGC CAA AAC Gly Asp Ile Lys Glu Tyr Gln Ala Leu Ala His Cys Phe Gly Gln Asn

	GGC Gly		GTC		TCC
	GAA Glu		AAG Lys	0	TCA
	GAC	1650	CTG	1700	AAC
009	CCG Pro	16	AAA Lys		CAT
П	GAC		GAG Glu		GGC Gly
	GAA Glu	01	AAG Lys	1690	GGC Gly
069	AAT TTG Asn Leu	1640	AAG Lys	<b>-</b>	TTC
1	AAT Asn		CCT		GGG G1y
	ATT Ile		GGC G1y	1680	TTT Phe
20	AAT Asn	1630	GTC	16	TCA
158	CCA AA		CTC		AAT Asn
	CAT His		CTG	0,	TCC
	ATC Ile	1620	AAA Lys	1670	TTG
1570	TGG Trp	16	GCA Ala		GGT Gly
-	GGA G1y		GAT Asp		GTC Val
	ACA Thr	1610	GTG Val	1660	AAG Lys

FIGURE 3 5 OF 6

GGA GGA GCT GGT GGC GTA GAA GCA GTT GCA GTA GTT CAG GCA ATA AGG Gly Gly Ala Gly Val Glu Ala Val Ala Val Glu Ala Val Ala Val Gln Ala Ile Arg

AGT GAG CTG AGA GTG AAT TCC ACC AAA TCG ATG ATC GGT CAC CTT CTT Ser Glu Leu Arg Val Asn Ser Thr Lys Ser Met Ile Gly His Leu Leu

1					
ATA CTA TTT Ile Leu Phe	GCC CCC TK Ala Pro C	GCC CCC TGC AAC TAG A AAAGAGTCTG TGGAAGCCGA GAGTCTTTGA Ala Pro Cys Asn ***	A AAAGAGTCTK	3 TGGAAGCCGA	. GAGTCTTTGA
1770	1780	1790	1800	1810	1820
TGC	ACGTTAGTAG	CTTCTTATGC	CTCTGAAACC	GAACTCATGC ACGTTAGTAG CTTCTTATGC CTCTGAAACC GAGATAGACC GGCTACTCGA	GGCTACTCGA
1830	1840	1850	1860	1870	1880
CCA	AAGATACTCC	TTGCCGGTAT	* TGGTGTTAAG	* GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	TTGTCCCTTT
1890	1900	1910	1920	1930	1940
TTC	TTCTTTTGAG	AGCTTTAACC	GAGGTAGTCG	TATITICITC TICTITIGAG AGCTITAACC GAGGTAGTCG TATITICGAG CTTITCGAAT	CTTTTCGAAT
1950	1960	1970	1980	1990	2000
CGT	TATCGGATCA	ATGTGTTTCT	TCTAAGATCA	* ACATGTTCGT TATCGGATCA ATGTGTTTCT TCTAAGATCA TTTGTAATGC ATATTTTGAA	ATATTTTGAA
2010	2020	2030	2040		
ATC '	TCAGTATGCA	AAACCACATC TCAGTATGCA AAATAAAAAA AAAAAAAAAA	AAAAAAAA	AAAAA	

FIGURE 3 6 OF 6

Sequence Range: 1 to 1921

					·							
09	CTACACCTCC	120	GCTCAATCGA	180	GCTCTGCAAC CTGCACAGGA AGTTACCACA	ATG Met>		AAT Asn>		GAT TGT Asp Cys>	370	ACA Thr>
	CTAC		3CTC		AGTT2	GGA Gly		AAT Asn	320	GAT Asp	, W	TCC
20		110		170	3GA 7	220 GTG ACT Val Thr	270	TAC	.,	TTT Phe		TTC
	ATGA(		ACCG		CACA			TTC Phe		ACC		TCT Ser
	TTCGAGCCCT GCCATGACTA		ACCACCCGCA GGCACCGGAG		CTG(	GTT Val		GAT GTT Asp Val	310	GAG Glu	360	AAG Lys
40	CCCT	100	CGCA	. 160	CAAC	GTA Val	260	GAT Asp	3.	ATA Ile	•	ATC Ile
	CGAG		CACC	•	rc <sub>TG</sub>	210 CGA Arg		CCT Pro		GAG Glu		GAG Glu
	TT	0	C AC	0		CGG Arg		GAC Asp		ATA AGT Ile Ser	350	GCT GGA Ala Gly
30	CTG	90	rccg	150	TGT	CAG Gln	250	CAT	300		(*)	GCT
	CGGCACGAGG TCACCTCTTA CCTCGCCTGC		TCGGATCCAG GCCCATCCGC		GCTTCCCCTT CCGGGGAGGC AATGGCTGTG	200 ATC AAA CAG Ile Lys Gln	25	GGC CAT (Gly His )		GGC G1y		ACG AGA ATT Thr Arg Ile
20	F. C.	80	}G G(	140	3C A2	ATC Ile		CTA		ACG AGT Thr Ser	340	AGA Arg
.,	TCT	w	ATCC?	16	GAGG	AGT Ser		CCT	290	ACG Thr	34	
	rcaco		rcGG2		3000	CCA Pro	240	ACT Thr	(1	GGA Gly		CCT
10	AGG 1	7.0	rgt 1	130	TT			GTG Val		GAT Asp		TTT Phe
	CACG2		GCATCCTTGT		יככככ	AAG Lys		GTG Val	30	CTT Leu	330	CAA Gln
	)550		GCA		GCTJ	AAG Lys	230	GGT Gly	280	CTG		GCT

FIGURE 4

420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	610	TTC Phe>	099	TGG Trp>	TTT Phe>
	TTC	GGA Gly		GTT Val	260	ATT Ile	61	CCT		GGA Gly	AAC Asn
	AAG Lys	460 AAT GGT Asn Gly	510	GGA Gly	٠,	GCC		GTA Val		TTG	700 ACG AGT Thr Ser
410	GAC Asp			TGC		GAT Asp		TGT Cys	650	GAC Asp	
•	ATG Met	ACA Thr		AAA Lys	550	AAT Asn	<b>*</b>	TTT Phe	v	ATG Met	GCA Ala
	AGG Arg	TTA Leu	200	AGA	5.	TTC Phe		CCC		GCA Ala	TGT Cys
400	TCC AAG Ser Lys	450 GCA Ala	Δ,	AAA Lys		GTA Val		AAT Asn	640	CTT Leu	690 GCT Ala
4	TCC	AAA Lys		GAT Asp		AAG Lys	290	ATG Met	9	ATG Met	ACT
	CTC	AAG Lys	490	CTA Leu	540	ATG Met	u,	AAG Lys		GCT Ala	TCT Ser
	AAG Lys	440 GCC GGC Ala Gly	4.9	GAG Glu		GGA Gly		AAG Lys		TCA	680 ATA Ile
390	CCG			AAA Lys		GGT Gly	580	$\mathtt{TAT}$	630	GGA Gly	TCG
	GCC Ala	ACT		ATG Met	530	ATG Met	28	TCA		ATG Met	TAC
	GTG Val	430 G CTG t Leu	480	GTG Val	٥,	GCA Ala		ATT Ile		AAT Asn	670 CCC AAC Pro Asn
380	TGG Trp	43 ATG Met		GAT Asp		TCA		AGG Arg	620	ACA Thr	670 CCC AJ Pro As
	GGT Gly	TAC		GAA Glu	520	GGC	570	CTA Leu	v	ACC Thr	GGC Gly
	GAT Asp	CTT	470	ACC	57	ATT Ile		GCC Ala		GCT	ATG

FIGURE 4 2/6

	GTG Val>		GGA Gly>	0.0	ACT Thr>	* 006	GGG G1y>	AAA Lys>		TGC Cys>		ATT Ile>
	GAT Asp	800	ATG Met	85	CCT Pro		ATG Met	AAG Lys		ACT Thr	1040	GTG Val
750	GCA Ala	~	GGT		GAC Asp		GTT Val	940 T GCA s Ala	066	TTC	. 1(	GGA G1y
	GAA Glu		ATT Ile		GCC Ala	890	TTT Phe	9, CAT His		AGT		GCT Ala
	GGC Gly	190	CCT Pro	840	AAT Asn	w	GGA Gly	GAG Glu		GGA Gly	30	GAT GGA Asp Gly
740	AGA Arg	7.	ATA Ile		AGA Arg		GAT Asp	TTA Leu	980	GGT Gly	1030	GAT Asp
•	ATC Ile		ATC Ile		CAG Gln	880	AAT CGT Asn Arg	930 GAG Glu	O,	CTA Leu		CCT
	ATA Ile		GTA Val	830	TCA	õ		GAG Glu		TTT Phe		CAC His
730	CAC His	780	GCG	~	TTG		AGT	CTA	970	GAA Glu	1020	CCT Pro
7	AAC Asn		GAT Asp		GCT		GAC	920 CTA	6	GCA	` '	GAG Glu
	GCG Ala		TCA	820	CGA Arg	870	TGG Trp	CTA		TAC		ACC Thr
	GCT	170	GGC Gly	88	TGC		CCA	GTG Val		ATT Ile	1010	CAC ATG His Met
720	AAT Asn	•	GGG G1y		GCA Ala		TCA AGA Ser Arg	910 GCT GGA Ala Gly	* 096	ACT Thr	1(	CAC His
	CTG		TGC		GTT Val	860		91 GCT Ala		GCG Ala		TAC Tyr
	ATC Ile	092	CTT	810	TTT Phe	~	GCT	GGA Gly		GGT Gly	00	GCC Ala
710	TGT	7	ATG Met		GGT Gly		AAA Lys	GAA	950	AGA Arg	1000	GAT Asp

FIGURE 4

06	GAC Asp>	1140	ATC Ile>	TTA Leu>		GCC Ala>		TGG Trp>	0	ACC Thr>	1380	GGT Gly>
1090	GAA Glu	-	GAT Asp	GAG Glu		GCA	1280	$\frac{\text{GGG}}{\text{Gl} Y}$	1330	GAT ACC ASP Thr	П	GTC Val
	AGG Arg		GGA Gly	1180 AAC AAC Asn Asn	1230	$_{ m GGA}$	12	ACT		GTG Val		AAG Lys
	TCT Ser	1130	GCT	1180 AAC AZ ASD AS	*-1	CTC		AGG Arg		GGC Gly	1370	AAC ATT Asn Ile
1080	GTC Val	H	CCA	CAA Gln		CTT	07	ATA	1320	GAT GAA GGC Asp Glu Gly	13	
•	$_{\rm GGA}$		ACT Thr	GGC G1y	1220	CAC His	1270	GCA Ala	-	GAT Asp		CTG
	TCA	50	TCC	1170 TGT TTC Cys Phe	ï	$_{\rm GGT}$		CAG Gln		CCA	0.9	AGA Arg
1070	CAG Gln	1120	ACA Thr	TGT		TCA ATG ATT Ser Met Ile		GTT Val	1310	GAA AAC ( Glu Asn 1	1360	GAG Glu
Ä	GCT		GCC Ala	CAC	10	ATG Met	1260	TCA GTA Ser Val	7	GAA Glu		AAG Lys
	TTG		CAT His	1160 CTT ATC Leu Ile	1210	TCA		TCA		TTG		AAG Lys
09	AAG GCT Lys Ala	1110	GCA Ala			AAA Lys		GTT Val	00	AAT Asn	1350	CCT Pro
1060		• •	AAT Asn	GCT		ACC Thr	1250	GCA Ala	1300	ATT Ile	П	GGC Gly
	GAG Glu		ATA Ile	1150 TAC CAA TYr Gln	1200	TCT Ser	12	GAA Glu		AAT Asn		GTG Val
	ATA Ile	1100	TAC Tyr	1150 TAC C TYF G	•	AAT Asn		GTG Val		CCG Pro	1340	CTC
1050	TGC	÷	AAT Asn	GAG Glu		GTG Val	0.1	GGT	1290	CAT His	13	TTG
••	CTC		GTA Val	AAA Lys	1190	AAA Lys	1240	$_{\rm G1Y}^{\rm GGT}$	1	ATC Ile		AAA Lys

FIGURE 4

CTC TTC Leu Phe>	1480	CAAA	1540	ATGCCCATG	1600	GCGACACAG	1660	TTCTGAAAT	1720	AAGAGAACA	1780	TTTATCGCCG	1840	TCATTGGAG
1420 AAC TCG TCC ATA Asn Ser Ser Ile	1470	CATGTGGA ATTCTACTCA ATCTATCAAA	1530	GCTGAAGTTT TGAGGACTCC AGCATGTTGG TAGCTCCTTA CGTCTCTAGA CATGCCCATG	1590	CGGGAGCTGT AGTCGGAACC ATGACGGATT GAGTACTCAT GGCGACACAG	1650	CTATTCATTA TCCCATTTTT TTTCTGAAAT	1710	CTCCCTCCTT ACGGTAGTTG TACTTTCGAG CGTTTCATCG AGTCAGTGAA GAAGAGAACA	1770		1830	TTTTGIGGGF TAAAATTTGT AAAACTAGAC GACTGGTTTG TTTTCTCTTG ATCATTGGAG
1410 GGT GGG CAC AAC Gly Gly His Asn	1460	GTGGA ATTCT	1520	TAGCTCCTTA	1580	ATGACGGATT	1640	CTATTCATTA	1700	CGTTTCATCG	1760	CCCTTTGTTT TGCTCTCTAT	1820	GACTGGTTTG
1400 GGG TTT GGT G Gly Phe Gly G	1450	TAG GGCGTTT CATGI ***>	1510	AGCATGTTGG	1570	AGTCGGAACC	1630	TGTTAGAGCA	1690	TACTTTCGAG	1750	TAACCATTTG	1810	AAAACTAGAC
TCA TTC Ser Phe	1440	AAC Asn	1500	TGAGGACTCC	1560	CGGGAGCTGT	1620	GATATACTCC TTGCTAGAAT	1680	ACGGTAGTTG	1740	GGGCACGTAG	1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC Ala Pro Tyr	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CICCCICCII	1730	AAGCTAACTC	1790	TTTTGTGGGT

FIGURE 4

1900	AAAAAAAA		
1890	ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAAA AAAAAAAAA		
1880	ATAAAAAAA		
1870	TTCATTGATG		6
1860	ATATTTGCCT	1920	****
1850	ATGTATGGCC	1910	A A A A A A A A A A A A A A A A A A A

FIGURE 4 6/6

	457	409	361	313	265	217	169	120	09
CGCCGCC ATG CAT TCC CTC CAG TCA CCC ASP His Ser Leu Gln Ser Pro 15 Ser Ileu Gln Ser Thr 15 Ser Ile Pro Arg Pro Lys Ser Ser Thr 15 Ser Ile Pro Asn Val Arg Ala Ala Ser Ser Ile Pro Asn Val Arg Arg CGC GCT TCC Arg Glu Thr Asp Pro Lys Lys Arg Val Ser Val Phe Gly Ser Asp Val Asp Val Ser Val Phe Gly Ser Asp Val Asp Val Ser Gly Glu Ser Gly Ile Gly Pro Ile Ser Gly Glu Ser Gly Ile Gly Pro Ile Ser Gly Glu Ser Gly Ile Gly Gly Glu Ile 95 TTC CCC ACG ACG GCC CAG ATT Phe Pro Thr Arg Phe Gly Gly Gly Gln Ile 95 TAC ATT GCC AAC AAC GCC CAG ATT Phe Pro Thr Arg Phe Gly Gly Gly Gly Ile 95 TAC TAC ATT GCC AAC AAC AAC AAC AAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC GCC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC AAA AAC GAC AGG CGC TAC ATT TAC ATT GAC AAA AAC GAC AGG CGG TAC ATT TAC ATT GAC AAA AAC GAC AGG CGG TAC ATT TAC ATT GAC AAA AAC GAC AGG CGG TAC ATT TAC ATT GAC AAA AAC GAC AGG CGG CGC CGC CGC CGC	Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser 80  CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG Pro Thr Arg Phe Gly Gln Ile Arg Gly Phe Asn Ser Met 100  ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC Ile Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Ile Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg 125	GGC GAG AGC GGG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser 80	TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu 65	GAG ACC GAC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly 50	ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro 45	CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg 20	ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro 1	TTCTTCTTCC ACCGCATCTC TTCTCTTCTC TTGGCTTCTC	CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCACGCGT CCGTCTTCCC

FIGURE 5

553	601	649	697	745	793	841	888
GCC Ala	666 61y	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	GGC Gly
GGT Gly	GTT Val	TCT Ser	TAT Tyr	CTG Leu 205	TAC Tyr	CTT Leu	GGA G1y
CTC Leu 140	CTG	CAA	CCC Pro	GGT Gly	AAC Asn 220	GAT Asp	TTG
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC Leu	TCC Ser	GCT Ala 235	GGG G1y
GCC Ala	GGA Gly	GGG G1Y 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT Ile 250
GAC Asp	GCC Ala	GAC Asp	TTC Phe 185	ATT Ile	GCC Ala	GGT Gly	CCA Pro
GAG Glu	aga Arg	TCT Ser	CCT	GCT Ala 200	TGT Cys	CGT Arg	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC Arg	ATC Ile
TCT Ser	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile 230	GCA Ala
AAG Lys	GAC Asp	ACT Thr 165	AAA Lys	GCC Ala	TCC	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG Leu	CGG Arg 180	TCT Ser	ATT Ile	AAT Asn	GAG Glu
GGG G1y	AAG Lys	GGT Gly	CAC His	GGG G1y 195	TCA	GCT Ala	ACT Thr
GCC Ala 130	TCC Ser	GGT Gly	GGT Gly	ATG Met	<b>TAT</b> <b>TYT</b> 210	GCT Ala	66C 61y
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC Arg	GGA G1Y 160	GAG Glu	ACA Thr	CCA	CAT His	GCT Ala 240
TGC Cys	GAC ASP	ACA Thr	ATC Ile 175	ATT Ile	66C 61y	TTC	ATT Ile

FIGURE 5

937	985	1033	1081	1129	1176	1224	1272
ACT Thr 270	GAA Glu	CGA Arg	GAT Asp	TCT	GTC Val 350	GCC	AAA Lys
CAG Gln	GGT G1y 285	AAA Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	GAT Asp	GAT ASP 380
GAC Asp	GTG Val	GCA Ala	ATC Ile 315	$_{\rm GLY}^{\rm GGT}$	CCT	666 61y	AAG Lys
GAT Asp	TTT Phe	CAT His	GCA Ala	CTC Leu 330	TCA Ser	GCT	ACA
AAC Asn 265	GGT Gly	GAA Glu	GGT Gly	GGT Gly	GTC Val 345	CTA	AAC
AGG Arg	GAT Asp 280	TTG	GGA Gly	GAT	GGC Gly	ACT Thr 360	AAG Lys
CAA Gln	CGT Arg	AGC Ser 295	TTG Leu	GCT	GCT Ala	TCT Ser	TTC <b>Phe</b> 375
TCT	GAC Asp	GAG Glu	TAT TY 310	AGG Arg	GAT	ACT Thr	GTT Val
CTG	AAA Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT Asp	GTG Val	GCA Ala	GAC ASP	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG Leu	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC Pro	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG Arg	GGA Gly	CCT Pro 305	CAC His	GAG Glu	ATA Ile	AAT Asn
GTG Val	TCT Ser	GCT Ala	GCA Ala	TAT Tyr 320	ATT Ile	TAC	ATA Ile
TTT Phe 255	GCC Ala	GGT Gly	${\tt GGA} \\ {\tt G1} Y$	GCT	TGC Cys 335	AAT Asn	GAG Glu

FIGURE 5

ATT AAT GCA ACT AAG TCA ATG ATC GGA CAC TGT CTT GGA GCC TCT GGA 1320 385 386 390 390 395 390 395 390 395 390 395 390 395 390 395 390 390 395 390 390 390 390 390 390 390 390 390 390
ATT Ile GGT GIY His AAT AAT AAT AAT TTC Phe TCAA

FIGURE 5

Sequence Range: 1 to 1802

09	TTATCTCCGC	.0 CCT TCC Pro Ser		TCC Ser	210	CGT Arg		CGG Arg		GTC Val	CTA
	rtatc	110 TCC CCT Ser Pro	160	Ser	7	ATC Ile		AAG Lys		GAC Asp	350 C GGC ATC AGC C r Gly Ile Ser I
20	rcg :	CAC 7 His 9		CCC		GTC Val		AAG Lys	300	TCC	35 ATC Ile
	CTTTCCGACC ACATTTCATT TCTTGCCTCG	CTC ( Leu 1		CTC AAT TCC Leu Asn Ser	200	CCC	250	CCC	•	66C 61y	GGC Gly
	TCT	100 TCC Ser	150	AAT Asn	7	CTC		GAC		T. 유	AG Se
40	CATT	CAA Gln		CTC		AGC		TCC	290	GTC Val	340 GAG Glu
	ATTT	ATG	•	CGC		GCC	240	GAG Glu	8	TCC	66C 61y
0	C AC	υ O U	140	CCC TTC Pro Phe	190	CGC Arg		CGC Arg		GTC Val	TCC
30	CGAC	၁၅၁၁၅ 06	À			CGT Arg		AAG Lys		CTC	330 CTG CTC Leu Leu
	TTTC	70 80 90 CGCTCCTCCG CCGTCGTTCG CCGCCGCCGC		GAG Glu		CTC	230	CCC Pro	280	GGC Gly	
20	3G C	08 090 090		CTC	180	CCC	ζ,	GCC Ala		ATG Met	AAG Lys
	rccG	CGTT	130	CCT	``	CGC		TCC		GGC G1y	320 TAC GAC AAG Tyr ASP Lys
	GGTCGACCCA CGCGTCCGGG	CCGT		TCC		CTC		GCC Ala	270	ACC Thr	
10	CCA (	70 20G		CCC	170	GCT Ala	220	ACC Thr	.,	ATC Ile	TAC
	:GAC	rccre	120	CGC Arg	H	GCC		GCC Ala		GTC Val	GCC
	GGTC	င္ပဗ္သ	<b>T</b>	CTC		GCC		GCT	260	GTC	310 GAC ASP

FIGURE 6 1/5

	CAG Gln	450	CGG Arg		GCT		AAG Lys	GTC Val		ATC	069	CTG	
400	GGC Gly	7	GAC Asp		AAG Lys		GAT Asp	ACT Thr	640	AAG Lys	v	GCG Ala	
	GCC		AAC Asn		AAG Lys	540	ATT Ile	590 CTA AC Leu Th		CGG Arg		TCT	
	TTC	440	AAG Lys	490	GGC Gly	•,	AAG Lys	GGC Gly		CAC His	089	GGG G1y	
390	AGG Arg	4	GGC		GCC		TCC	GGT Gly	630	$_{\rm GGT}^{\rm GGT}$	39	ATG Met	
.,	ACC Thr		GAC		GTC Val	530	CTC	580 ATG Met	Ψ	AAA Lys		AAC Asn	
	CCC		ATC Ile	480	ATT Ile	53	TCC	GGT Gly		GAG Glu		ACA Thr	ø
380	TTC	430	TAC	7	TGC		CAA Gln	ACC	620	ATC Ile	670	ATT Ile	URE /5
ñ	AAA Lys		GGC Gly		TAC		GGC Gly	570 GGA Gly	9	CTC		GCC	FIGURE 2/5
	TCC		ACG Thr	470	CGC Arg	520	GCC Ala	GTT Val		AAT Asn		TAT	
	GCT	420	GCG Ala	4.7	CTC		CTC	CTA		CAG Gln	* 099	CCA Pro	
370	GAC Asp	4.	AAC Asn		TGC		GAT Asp	50 GTG Val	610	GTT Val	Ψ	ATT Ile	
	TTC Phe		TTC Phe		GAT Asp	510	GCC	560 GGA G1 Gly Ve		$_{\rm GGG}$		TTC Phe	
	CGC Arg	410	GGC G1y	460	GAC	Δ,	GAC	GCC		GAC	650	TTT Phe	
360	GAC Asp	4.	CGT Arg		CTC		GAA Glu	AGG Arg	* 009	TCT	9	CCG	
. ,	ATC Ile		ATC Ile		CGG Arg	200	CTC	550 GAG Glu	v	TTC Phe		TCC	

SUBSTITUTE SHEET (RULE 26)

FIGURE 6 3/5

	ACT Thr		ATC Ile	GCG Ala		TCT	930	GAC		GAG Glu		TAT Tyr
	TCA		CAT His	GCT Ala	880	TTA Leu	0,	AAG Lys		ATG Met		GAA Glu
	ATT Ile	780	AAT Asn	830 GAG GCT Glu Ala		GCT Ala		GAT Asp		GTT	1020	GCA
730	TCG	1-	GCC	ACT Thr	·	AGG Arg	920	TGG Trp	970	TTG GTT Leu Val	10	ATT Ile
	TAT Tyr		GCC	GGA Gly	870	GCC TGC AGG Ala Cys Arg	92	CCG		GTA Val		ATT Ile
	AAC Asn	170	GCT	820 GGA Gly	ω	GCC		AGG Arg		GGA Gly	01	CCG
720	CCA AAC Pro Asn	7.7	TAT Tyr	GCT		GTT Val		TCA	096	GCT	1010	GCG Ala
	GGC Gly		TTT Phe	ATT Ile	098	TTC	910	GCC	O1	GGG Gly		GGA Gly
	ATG Met		TGC	810 ATG Met	8	GGA Gly		ACT Thr		GAA Glu		CGG Arg
710	CTG	760	TAC	CTG		GGA Gly		CAG Gln	950	$_{\tt GLY}^{\tt GGT}$	1000	AAA Lys
7.1	GGT Gly		AAC Asn	GAC Asp		TTA Leu	006	CCT	6	ATG GGT Met Gly	<b>T</b>	ATG Met
	TTG		TCC	800 G GCT u Ala	850	GGT Gly	O,	GAT Asp		GTG Val		GCA Ala
	GAT ASD	750	ACT Thr	80 GAG Glu		ATT Ile		GAT Asp		TTT Phe	066	CAT His
700	ATC Ile	•	GCT	GGT		CCA	890	AAT Asn	940	GGC Gly		GAG Glu
	GCC		TGT	CGA Arg	840	ATT Ile	8	AGG Arg		GAT Asp		TTG
	CTT	740	GCA	790 CGC Arg	<b>~</b>	GTC Val		CAA Gln		CGT Arg	980	AGC Ser

SUBSTITUTE SHEET (RULE 26)

AGG Arg		GAT Asp	1170	ACT		GTT Val		ATC Ile	ATT Ile		AAT
1070 GAT CCA ASP Pro	1120	GAA Glu	11	GCG		AAA Lys		ATG	1280 1310 CTT GGA GCA TCA GGA GGT CTT GAA GCC ATC GCA ACC Leu Gly Ala Ser Gly Gly Leu Glu Ala Ile Ala Thr	1360	AAG GGA ATA ACC ACC GGC TGG CTT CAT CCC AGC ATT AAT CAA TTT AAT Lys Gly Ile Thr Thr Gly Trp Leu His Pro Ser Ile Asn Gln Phe Asn
1070 GAT C( ASP P)	•••	CTC		CAT His		GCC ATT AAG Ala Ile Lys	1260	AAG TCA Lys Ser	131 GCA Ala		CAA
ACT		AGT Ser	09	AAT GCT Asn Ala	1210	ATT Ile	1.		ATC Ile		AAT
ATG Met	1110	AGC	1160	AAT	•	GCC Ala		ACT Thr	GCC	1350	ATT Ile
GGA GGT GCA GTC AAC TGT GAT GCT TAT CAT ATG ACT Gly Gly Ala Val Asn Cys Asp Ala Tyr His Met Thr	H	TCG TGC ATT GAG AGC AGT Ser Cys Ile Glu Ser Ser		ATA		AAT Asn	20	AAT GCA ACT Asn Ala Thr	1300 GAA Glu	H	AGC
TAT Tyr		ATT Ile		AAT TAC Asn Tyr	1200	GAG ATA Glu Ile	1250	AAT Asn	CTT		CCC
1050 TGT GAT GCT Cys Asp Ala	1100	TGC	1150	GTC AAT TAC Val Asn Tyr	1	GAG		GAA ATC AAA ATC Glu Ile Lys Ile	GGT Gly	40	CAT
050 GAT ASP	11	TCG				GCC		AAA Lys	290 GGA Gly	1340	CTT
1 TGT Cys		TCC		GAG Glu	1190	GAT CTT Asp Leu	1240	ATC Ile	1 TCA Ser		TGG
AAC		GGT GTC GIY Val	1140	CCT GAA Pro Glu	11	GAT			GCA Ala		GGC Glv
1040 GCA GTC Ala Val	1090	GGT	7			$\frac{\text{GGG}}{\text{G1}\text{y}}$		ACC AAG Thr Lys	80 GGA Gly	1330	ACC
10 GCA		CTT		TCA		GCT	1230	ACC Thr	12 CTT Leu		ACC
GGT		GGG	1130	GGG GTC Gly Val	1180	CTT	ਜ	AAC Asn	TGT Cys		ATA Ile
	1080	GCT GAT Ala Asp	11			ACT Thr		AAG Lys	CAC	1320	AAG GGA Lys Gly
1030 TTG Leu	Η	GCT		GCC		TCT	1220	TTC	1270 GGA Gly	ਜ	AAG

FIGURE 6

1410	CAG CAA Gln Gln		GGG CAC Gly His	1510	CTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	CTCTGATTTA	1750	STTATTTAAG		Ŧ.
1400	AAC ACT GTT GCC AAC AAA AAG Asn Thr Val Ala Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TTC TCA GCT TTC AAG CCA TGA ATTCT ACTTGGTTCA Phe Ser Ala Phe Lys Pro ***	1560	AAATGCACAC CAGTTGCTGA GATAGGGCTT CAACTTGCAG AGCAATTTTT TAAATGCCTT	1620	GAATAGGTCG GTCCTTTGAT AGTTCCTCGA AGCCATTTAG	1680		1740	TGTATTAGAA AGACCAATGA AAGATTTTGT GTCATGTTTG TGTTGTCAAT GTTATTTAAG	1800	ATAAAGCAAA AAAAAAAA AAGGGCGGCC GCTCTAGAGG ATCCAGCTTA CT
1390	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TC AAG CCA the Lys Pro	1550	CAACTTGCAG	. 1610	GTCCTTTGAT	1670	TACTGTAATA ATCGAAGATG ATTCCCATTT TAAATCTAGT	1730	GTCATGTTTG	1790	GCTCTAGAGG
	GTG GAC TTC AAC N Val Asp Phe Asn 1	1430	GCT ATC TCG A	1480	C TCA GCT 1 Te Ser Ala E	1540	GATAGGGCTT	1600	GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGGCC
1380	TCG		AAC GTC Asn Val	1470	GTT GTG GCA TT Val Val Ala Ph	1530	CAGTTGCTGA	1590	CGTAATACCG	1650	TACTGTAATA	1710	AGACCAATGA	1770	AAAAAAAAA
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	1460	AAC TCG GTJ Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

FIGURE 6 5/5

Sequence Range: 1 to 2369

	CATAAAAGAG	120	TTACCATACC	180	ATCCTTTTCT	230 GCC TCT TCC Ala Ser Ser>	280	Met Ser>	330	CTCT CCT		CCA CTA Pro Leu>
50	CACGCGTCCG	110	CTTCGATTCA	170	CCCAAAGGGT	GCC	270	GCC GCC TGC Ala Ala Cys	320	TCC ATC TCC Ser Ile Ser	370	CAA TGC GCC Gln Cys Ala
40	CGGGTCGACC	100	CTCCTTTCAT	160	GGTCTTTCAT	22 CCTCCA ATG Met	0	TGG CTC CTT Trp Leu Leu	310	CTT CCG CCT Leu Pro Pro	360	ATT CTC TCC Ile Leu Ser
30	CCGGAATTCC	06	TGCGGCCACC	150	GCCTTTTCCG	210 220 CAGTCAGTTC CCTCCA ATG CCT Met Pro	260	TGT ACG Cys Thr	0 *	GAC CCT Asp Pro	350	CGC CGG Arg Arg
20	GTACGCCTGC AGGTACCGGT CCGGAATTCC CGGGTCGACC CACGCGTCCG	80	AGAGAGAGG ATCCATCGAA TGCGGCCACC CTCCTTTCAT CTTCGATTCA	140	ATTCCGCTGA TCCATTTTCC GCCTTTTCCG GGTCTTTCAT CCCAAAGGGT ATCCTTTTCT	200 CTCAAAGGGT	250	TCC CCT CTC Ser Pro Leu	300	CAC CCC TCC His Pro Ser	340	CGC CTC TCC CGC Arg Leu Ser Arg
10	GTACGCCTGC	70	AGAGAGAGGG	130	ATTCCGCTGA	190 ATCCTATCTT	240	CTG CTC GCT Leu Leu Ala	290	ACC.TCC TTC Thr Ser Phe	3	CGC CGA CGC Arg Arg Arg

FIGURE 7

 $F_{\epsilon}$ 

	GTC Val>	TCC Ser>	0.	CGG Arg>	570	CTG Leu>		CAG Gln>		CAT His>	ATA Ile>
	CTC	470 ACA Thr	520	CAC His		GCT		AAA Lys		GGC Gly	710 AGT GGC Ser Gly
420	ACC	, TAT TYT		AGG		GTG Val	610	ATC Ile	* 099	CTA Leu	
	CAT	TAC		CGC		GCC	61	AGT Ser		CCT	ACG
	TTC	50 GAC Asp	510	ACC	u,	ATG Met		CCA		ACT	700 GAT GGA ASP Gly
410	AGT	460 C CAT GA S His AS		ACC		GCA Ala		AAG Lys	029	GTG Val	70 GAT ASP
•	TC( Se)	TGC Cys		CGC Arg	550	GAG Glu	009	AAG Lys	v	GTG Val	CTT Leu
	GGA Gly	CCC	200	CCC ATT (Pro Ile )	55	AGG		AAG Lys		GGT Gly	CTG
400	CGC	450 GAG Glu	-,	CCC		TCC		ACA	640	ATG Met	690 <b>AAT</b> Asn
4	CTC	TTC		AGA Arg		CCT	290	ACC	9	GGA Gly	AAT
	GCC	TGC	490	TCC	540	TCC	٥,	GTT Val		ACT	TAC
	TCC	440 GCC Ala	4.9	GGA Gly		GCT Ala		GAA Glu		GTG	680 TTC Phe
390	TCC	CTC Leu		TTC Phe		CGA Arg	580	CAG	630	GTT Val	6 GTT Val
	GCT	TAC Tyr		TTG	530	AAT Asn	28	GAA Glu		GTA Val	GAT
	TCT Ser	TCT Ser	480	TCC	וְאַ	CTC		CCT		CGA Arg	O CCT Pro
380	CCT	430 ACC TCT Thr Ser		GCA Ala		AGG Arg		CAA Gln	620	CGG	670 GAC CCT ASP Pro

1GURE 7

160	GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	00	AAG Lys>	1050	GCT Ala>
7(	ATT Ile		AAG Lys		GGC Gly		GAG Glu	950 GGA Gly	1000	AAG Lys		TCA
	AGA Arg		CCG	850	GCT	*	AAA Lys	S GGT Gly		TAT		GGA Gly
	ACG Thr	800	GCC	8	ACC Thr		ATG Met	ATG Met		TCA	1040	ATG Met
750	CCT	ω	GTG Val		CTG		GTG Val	940 TCA GCA Ser Ala	066	ATT Ile	10	AAT Asn
	TTT Phe		TGG Trp		ATG Met	890	GAT			AGG Arg		ACC ACA I
	CAA Gln	790	$_{\rm G1y}^{\rm GGT}$	840	TAC Tyr	ω	GAA	GGC Gly		CTA Leu	0	ACC Thr
740	GCT Ala	7.5	GAT Asp		CTA Leu		ACC Thr	ATT Ile	086	GCC	1030	GCT Ala
	TGT Cys		ACA Thr		ATG Met	880	ATC Ile	930 CTC Leu	O1	GAA Glu		TTC
	GAT Asp		TCC	830	TTC	88	GGA Gly	GTT Val		ATT Ile		CCT Pro
730	TTT Phe	780	TTC Phe	w	AAG Lys		$_{\rm GGT}^{\rm GGT}$	GGA Gly	970	GCC	1020	TGT GTA Cys Val
7.	ACC		TCT Ser		GAC		GAT Asp	920 TGC Cys	97	GAT Asp	-	
	GAG Glu		AAG Lys	820	AGG ATG Arg Met	870	ACA Thr	aaa Lys		AAT Asn		TTT Phe
	ATA Ile	170	ATC Ile	8	AGG Arg		TTA	AGA Arg		TTC Phe	1010	CCC
720	GAG Glu	• -	GAG Glu		AAG Lys		GCA Ala	910 GAT AAA ASP Lys	* 096	GTA Val	10	AAT Asn
	AGC		GGA Gly		TCT	860	AAA Lys	91 GAT ASP		AAG Lys		ATG

TGURE 7

FIGURE 7

	TCT Ser>		CAT His>	GCG Ala>	0	TTG Leu>	1290	AGT Ser>		CTA Leu>		GAA Glu>
	ATA Ile		AAC Asn	1190 TCA GAT Ser Asp	1240	GCT	П	GAC AGT Asp Ser		CTA Leu		GCA
0	TCG	1140	GCG			CGA		CCA TGG Pro Trp	30	CTA	1380	ATT TAC Ile Tyr
1090	TAC	•	GCT Ala	66C G1y		TGC	1280		1330	GTG Val		ATT Ile
•	AAC Asn		AAT Asn	1180 TGC GGG Cys Gly	1230	GCA Ala	17	AGA Arg		GGA Gly		ACT Thr
	CCC	1130	ATG	1180 TGC GC Cys G	-	GTT Val		AAA GCT TCA Lys Ala Ser		GCT Ala	1370	GCG Ala
1080	666 Gly	11	ATA Ile	CTT Leu		TTT Phe	. 07	GCT	1320	GGA Gly	H	AAA AGA GGT Lys Arg Gly
-	ATG		TGT	ATG Met	1220	GGT Gly	1270	AAA Lys	• •	GAA Glu		AGA Arg
	TGG Trp	0	TTT Phe	1170 GAT GTG Asp Val	17	GGA Gly		ACT Thr		$_{\rm GGG}$	20	AAA Lys
1070	GGA G	1120	AAC Asn			ATG Met		CCT	1310	ATG	1360	AAG Lys
10	$ ext{TTG}$		AGT	GCA Ala	01	GGT Gly	1260	GAC	ਜ	GTT		GCA
	GAC Asp		ACG Thr	1160 GGC GAA Gly Glu	1210	ATT Ile		TCC		TTT Phe		CAT His
0	ATG	1110	GCA			CCT Pro		AAT	00	GGA Gly	1350	GAG Glu
1060	GCA Ala	-	TGT Cys	AGA Arg		ATA	1250	aga Arg	1300	GAT Asp	•••	TTG
	CTT		GCT Ala	SO ATC Ile	1200	ATC Ile	13	CAG Gln		CGT		GAG Glu
	ATG Met	1100	ACT	1150 ATA ATC Ile Ile	П	GTA Val		TCC		AAT Asn	1340	GAG Glu

SUBSTITUTE SHEET (RULE 26)

CCT Pro>	1480	TTG GCT Leu Ala>	1530	GCC Ala>		CAC His>		ATG Met>	GTA Val>	1720	GAA Glu>
1430 C GAG r Glu	14	TTG		CAT His		ATC Ile		TCA	1670 TT TCA	17	TTG
1430 ACC GAG Thr Glu		GCT Ala		GCC	0,	CTT Leu	1620	AAA Lys	16 GTT Val		AAT Asn
ATG Met		AAG Lys	1520	AAT Asn	1570	GCT CTT Ala Leu		ACC Thr	GCA Ala		ATT AAT TTG Ile Asn Leu
0 CAC His	1470	GAG Glu	15	ATA Ile		CAA Gln		TCA	50 GAA Glu	1710	CCG AAT Pro Asn
1420 TAC CAC Tyr His	т	ATA Ile		TAC Tyr		$\mathtt{TAC}$	1610	AAT Asn	1660 GTG GA Val G1	<b>F</b> 1	CCG
SCC 11a		TGC Cys	0	AAT Asn	1560	GAG Glu	16	GTT AAT 1 Val Asn 9	GGT Gly		CAT His
1410 ACT TGC GAT C Thr Cys Asp A	1460	CTC	1510	GTA Val	П	AAA Lys		CAA AAC AGA GAG TTA AAA Gln Asn Arg Glu Leu Lys	GGT G1y	1700	TGG ATC Trp Ile
410 TGC Cys	14	ATT Ile		GAC Asp		GGA GAT ATC Gly Asp Ile	0	TTA	1650 GCA GCC Ala Ala	17	TGG
ACT Thr		GTG Val		GAA Glu	1550	GAT Asp	1600	GAG Glu			666 Gly
rTC Phe	0	GGA Gly	1500	AGG	15			AGA Arg	GGA Gly	0	ACT Thr
1400 GGG AGT G Gly Ser	1450	GCT	7	TCT		GCT		AAC Asn	1640 CTT CTC Leu Leu	1690	ATA AGG ACT Ile Arg Thr
14 GGG Gly		GGA Gly		GTC Val	0	CCG Pro	1590	CAA	16 CTT Leu		ATA Ile
GGT Gly		GAT GGA Asp Gly	1490	GGA Gly	1540	ACT Thr	П	GGC Gly	CAC		GCA
O CTA Leu	1440	CCT	14	TCA		TCC		TTC Phe	1630 ATT GGT Ile Gly	1680	CAG Gln
1390 TTT CTA Phe Leu	7	CAC		CAG		ACA	1580	$^{ m TGT}$	1630 ATT G Ile G	-	GTT Val

FIGURE 7 5/7

1770	AAG AAG Lys Lys>		TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920	ACTCCAGCAT	1980	GCTTTAGTCG	2040	ACTCCTTGCT AGAATTGTTG	2100	CCTTGCAATA	2160	AACAAAGCTG TTAACTCGGG
1750	AAA TTG CTC GTG Lys Leu Leu Val	1800	TTG TCT AAT Leu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	TGTGTCCGGA GCTTTAGTCG	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	
	GAT ACA Asp Thr	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	TCAAAGCTGA	1960	CCATGAGTTT	2020	CACTTGATAT	2080	TTTTTCTCTG	2140	TGAAGAAGAG
1740	GAA GGC GTG Glu Gly Val	1780	CTG AAC GTT AAG Leu Asn Val Lys	1830	TCG TCC ATA Ser Ser Ile	1890	GIGGAATICI ACICAACAIA ICAAAGCIGA AGIIITIGAGG ACICCAGCAI	1950	CTAGACATGC	2010	CTCATGGCGA	2070	TCATATITIT TITITCTCTG AAAICTCCCT CCTTGCAAIA GITGTACTIT	2130	ATCGAGTCAG TGAAGAGAG
1730	AAC CCA GAT GAA Asn Pro Asp Glu	17	GAG AGA CTG Glu Arg Leu	1820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT	1940.	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

FIGURE / 6/7

AAAAAAAA	AAAAAAAAA	AAAAAAAAA	тесваратара дарадара дарадарара дарадарара дарадара	AAAAAAAA	TGGAAATAAA
2350	2340	2330	2320	2310	2300
ATGTATGTTT	TAATTGGGGR	TTCTCATTGA	AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	CTGGTTTAGA	AACTAGAAGA
2290	2280	2270	2260	2250	2240
AAATTTGTAA	TGTGGTTTTA	ATCACCGTTT	CCATTTGCCC TTTGTTTTGC TCTCTATTTC ATCACCGTTT TGTGGTTTTA AAATTTGTAA	TTTGTTTTGC	CCATTTGCCC
2230	2220	2210	2200	2190	2180

FIGURE 7

Sequence Range: 1 to 2374

09*	CACCAAAC	120	AGACAGAC	180	TTCGATTC	240	CCAAAGGG	300	TGCCGCCT	360	TCTACCTCCT	420	CICICCCGCC	480	SCGGATCCA
20	CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC CACACCAAAC	110	TCTCTTCTCA AGACGGACGC CATTGGCAGC AGACAGACAG ACAGACAGAC	170	CCTCCTTTCA TCTTCGATTC	230	GGGTCTTTCA TCCCAAAGGG	290	TATCCTATCT TCTCAAAGG TCAGTCAGTT CCCTCCAATG CCTGCCGCCT	350	CGCCTGCATG TO	410	TCGCCGACGC C	470	GCCGGATTCT CTCCCAATGC GCCCCACTAC CTTCTGCTTC CTCCGCCCTC CGCGGATCCA
40	ACGCGTCCGC	100	CATTGGCAGC	.160	ATGCGGCCAC	220	CGCCTTTTCC	280	TCAGTCAGTT	340	GGCTCCTTGC	400	CCGCCTTCCA TCTCCTCTCC TCGCCGACGC	460	CTTCTGCTTC
30	GGGTCGACCC	06	AGACGGACGC	150	GATCCATCGA	210		270	TCTCAAAGGG	330	CTCTGTACGT GGCTCCTTGC	390	CCGCCTTCCA	450	GCCCCACTAC
20	CGGAATTCCC	80	TCTCTTCTCA	140	CCATAAAAGA GAGAGAGG GATCCATCGA ATGCGGCCAÖ	200	CATTCCGCTG ATCCATTTTC	260	TATCCTATCT	320	CTICCCIGCI CGCTICCCCI	380	CGACCCTCTT	440	CTCCCAATGC
10	-A-CNTGGTC	70	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCTC	430	GCCGGATTCT

FIGURE 8 1/5

550 560 570 580 590 600 ** CATCGGCATC CTTGTTCGGA TCCAGACCCA TTCGCACCAC CCGCAGGCAC CGGAGGCTCA	SCAC CGGAGGCTCA		099 059	GGAGGCAATG GCCGTGGCTC TGCAACCTGA ACAGGAAGTT	710 720	TTGTGACTGG AATGGGTGTG	770 780	CTTG ATGGAACGAG	830 840	AGAA TTGCTGGAGA	006 068	CTCTCTAAGA GGATGGACAA	950 960	CTGCTGGCAA GAAAGCATTA ACAGATGGTG GAATCACCGA	1010 1020	AGATGTGATG AAAGAGCTAG ATAAAAGAAA ATGCGGAGTT CTCATTGGCT CAGCAATGGG
		CCGCAG		TGCAAC		TTGTGA		AATCTG		CCTACGAGAA		CTCTCT		ACAGAT		CTCATT
	280	TTCGCACCAC	640	GCCGTGGCTC	700	CGGCGAGTAG	760	TTTCTACAAT AATCTGCTTG	820	CCTTTGATTG TGCTCAATTT	880	GATCAAGTCT TTCTCCACAG ATGGTTGGGT GGCCCCGAAG	940	GAAAGCATTA	1000	ATGCGGAGTT
)	570	TCCAGACCCA	630	GGAGGCAATG	069	TATCAAACAG	750	TAGGCCATGA ACCTGATGTT	810		870	ATGGTTGGGT	930	CTGCTGGCAA	066	ATAAAAGAAA
	260	CTTGTTCGGA	620	CCCTTCCAGG	089	ACCACAAAGA AGAAGCCAAG TATCAAACAG	740		800	GAGATAGAGA	860	TTCTCCACAG	920	GTTCATGCTA TACATGCTGA	086	AAAGAGCTAG
opination pour point outrooting	550	CATCCGCATC	610	ATCGAGCTTC	019	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

FIGURE 8 2/5

ACCACATGAC	TGCGATGCCT	GAGTTTCACT	TTCTAGGTGG	TGCGACTATT TACGCAGAAT TTCTAGGTGG GAGTTTCACT TGCGATGCCT ACCACATGAC	TGCGACTATT
1500	1490	1480	1470	1460	1450
AGAAAAGAGG	GAGCATGCAA	AGAGGAGTTG	TGCTACTACT	TATGGGGGAA GGAGCTGGAG TGCTACTACT AGAGGAGTTG GAGCATGCAA AGAAAAGAGG	TATGGGGGAA
1440	1430	1420	1410	1400	1390
ATGGATTTGT	AGTAATCGTG	ACCATGGGAC	AAGCTTCAAG	GAGAAATTCC GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG ATGGATTTGT	GAGAAATTCC
1380	1370	1360	1350	1340	1330
CTTTGTCCCA	TIGGIAIGGG AGGITITIGIT GCAIGCCGAG CITIGICCCA	AGGTTTTGTT		AGATGCGGTA ATCATACCTA	AGATGCGGTA
1320	1310	1300	1290	1280	1270
GCGGGGCTC	GTGATGCTTT GCGGGGGCTC	CGAAGCAGAT	GCGAACCATA TAATCAGAGG CGAAGCAGAT	GCGAACCATA	AATGAATGCT
1260	1250	1240	1230	1220	1210
ACTTTTGTAT	GCAACGAGTA ACTTTTGTAT	TACTGCTTGT	ACTCGATATC	GGGATGGATG GGGCCCAACT ACTCGATATC TACTGCTTGT	GGGATGGATG
1200	1190	1180	1170	1160	1150
CAATGGACTT	GCTATGCTTG	TATGGGATCA	CTACCACAAA	TCCCTTTTGT GTACCTTTCG CTACCACAAA TATGGGATCA GCTATGCTTG	TCCCTTTTGT
1140	1130	1120	1110	1100	1090
AGAAGATGAA	АТТТСАТАТА	AGCCCTAAGG	ATGCCATTGA	TGGAATGAAG GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA AGAAGATGAA	TGGAATGAAG
1080	1070	1060	1050	1040	1030

FIGURE 8 3/5

TTTGTGTCCG	GCCCATGAGT	CTCTAGACAT	CTCCTTACGT	GGACTCCAGC ATGTTGGTAG CTCCTTACGT CTCTAGACAT GCCCATGAGT TTTGTGTCCG	GGACTCCAGC
2040	2030	2020	2010	2000	1990
GAAGTTTTGA	TATCAAAGCT	CTACTCAACA	GACGTTTCGT GTGTGGAATT CTACTCAACA TATCAAAGCT GAAGTTTTGA	GACGTTTCGT	TTACATCTAG
1980	1970	1960	1950	1940	1930
TCTTCGCCCC	TCGTCCATAC	TGGGCACAAC	TCTAATTCAT TTGGGTTTTGG TGGGCACAAC TCGTCCATAC	TCTAATTCAT	GGTCGGTTTG
1920	1910	1900	1890	1880	1870
TGAACGTTAA	AAGGAGAGAC	GGGTCCTAAG	GTGGATACAA AATTGCTCGT GGGTCCTAAG AAGGAGAGAC TGAACGTTAA	GTGGATACAA	AGATGAAGGC
1860	1850	1840	1830	1820	1810
TGGAAAACCC	AATATTAATT	GATCCATCCG AATATTAATT TGGAAAACCC	CAGGCAATAA GGACTGGGTG		TTCAGTAGTT
1800	1790	1780	1770	1760	1750
TGGAAGCAGT	GCCGGTGGTG	TCTCGGAGCA	TTGGTCACCT	TAATTCAACC AAATCAATGA TTGGTCACCT TCTCGGAGCA GCCGGTGGTG TGGAAGCAGT	AATTCAACC
1740	1730	1720	1710	1700	1690
AGTTAAAAGT	CAAAACAGAG	CTGTTTCGGC	стсттатсса	AGATATCAAA GAGTACCAAG CTCTTATCCA CTGTTTCGGC CAAAACAGAG AGTTAAAAGT	SATATCAAA
1680	1670	1660	1650	1640	1630
CTCCGGCTGG	GCCACATCCA	AAATGCCCAT	TAAATTACAT	AGGAGICTCT AGGGAAGACG TAAATTACAT AAATGCCCAT GCCACATCCA CTCCGGCTGG	SGAGTCTCT
1620	1610	1600	1590	1580	1570
TGGCTCAGTC	GAGAAGGCTT	TCTCTGCATA GAGAAGGCTT TGGCTCAGTC	CTGGAGTGAT	CCTGATGGAG	CGAGCCTCAC
1560	1550	1540	1530	1520	1510

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		ATCC	2370 GCTCTAGAGG	2350 2350 2340 AAAAAAAA AAGGCCGCCC GCTCTAGAGG ATCC	2350 AAAAAAAAA
TTTTCTCAAA	GATTGGTTTG	GACTGGTTTA	TAAAATTTGT AAAACTAGAA GACTGGTTTA GATTGGTTTG	TAAAATTTGT	TTTGTGGTTT
2340	2330	2320	2310	2300	2290
TCATCACCGT	CCTTTGTTTT GCTCTCTATT TCATCACCGT		GGCACGTAGT AACCATTTGC		TGTTAACTCG
2280	2270	2260	2250	2240	2230
TCATCGAGTC AGTGAAGAAG AGAACAAAGC	AGTGAAGAAG	TCATCGAGTC	TTCGAGCTTT	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT	CTCCTTGCAA
2220	2210	2200	2190	2180	.2170
TCTCATATTT TTTTTTCTC TGAAATCTCC	TTTTTTCTC	TCTCATATTT	TGGTAGAGCA ATATTCATTA		CTAGAATTGT
2160	2150	2140	2130	2120	2110
TACTCATGGC GACACTTGAT ATACTCCTTG	GACACTTGAT	TACTCATGGC	CGGAACCATG ACGGATTGAG		GAGCTTTAGT
2100	2090	2080	2070	2060	2050

FIGURE 8 5/5

Sequence Range: 1 to 1580

00	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	01	CGC Arg>
1(							GGA Gly	gor GCT Ala	Ä	ATC Ile
	CAG Gln			0		240	ATT Ile	CTT Leu		GGG Gly
	ACT Thr	40	TCC	13	TCT		TTA	GAT Asp		ACG
90	GCA Ala	П	GTC Val		CAG Gln		AAA Lys	i0 GAT ASP	330	CGA Arg
	AGG Arg		TTT Phe		AGG Arg	3.0	TGC Cys	28 AAT Asn		GTC Val
	AGA Arg	0	GAG Glu	180	GAC Asp	0	GGA Gly	TCA		ACT
80	CTG	13	TCG		TCT Ser		AGA Arg	GTC Val	20	ATT Ile
	GCC Ala		TCC		GAT	0	AGT Ser	270 CAA Gln	(C)	TGG Trp
	CCT		TCT	.70	CAG Gln	22	GTG Val	CTT Leu		GAA Glu
0	GTT Val	120	${\tt GGA} \\ {\tt G1y}$	П	GTT Val		CTT Leu	GCT Ala	0	
	TCA		CGT Arg		GCC		AGG Arg	CCA Pro	31	AAT GAT Asn Asp
	TCT		TCT Ser	0	AGT Ser	210	CCG Pro	2 ATA Ile		ACC
	$_{\rm G1y}^{\rm GGT}$	10	TCG Ser	16	TGT Cys		TCG	GCT Ala		GAC
<b>6</b> 09	CTG	П	TCA		TGC Cys		CGC	0 TCT Ser	300	GTC
	TTT Phe		ATT Ile		TTT Phe	200	TCT Ser	25 GGT Gly		ATT Ile
	70 80	60 70 80 90 10 * CTG GGT TCT TCA GTT CCT GCC CTG AGA AGG GCA ACT CAG CAT Leu Gly Ser Ser Val Pro Ala Leu Arg Arg Ala Thr Gln His	60 70 80 90 10  *CTG GGT TCT TCA GTT CCT GCC CTG AGA AGG GCA ACT CAG CAT Leu Gly Ser Ser Val Pro Ala Leu Arg Arg Ala Thr Gln His  110 120 130 140	60	Fig. 70 80 90 10  CTG GGT TCT TCA GTT CCT GCC CTG AGA AGG GCA ACT CAG CAT  Leu Gly Ser Ser Val Pro Ala Leu Arg Arg Ala Thr Gln His  110 120 130 140  TCA TCG TCT CGT GGA TCT TCC TCG GAG TTT GTC TCC AAA AGG Ser Ser Ser Ser Ser Glu Phe Val Ser Lys Arg  160 170 180 190 190	60         70         80         90         10           CTG         GGT         TCT         GCT         GCC         CTG         AGG         GCA         ACT         CAG         CAT           Leu         Gly         Ser         Val         Pro         Ala         Leu         Arg         ACT         CAG         CAG         CAG         ALA         Arg         ACT         CAG         CAG         ACT         ARG         CAG         ACT         CAG         ACT         ACG         CAG         ACG         CAG         ACT         ACG         ACG	Fig.   Fig.	Fig.   Fig.	CTG   GGT   TCT   TCA   GTT   CCT   GCC   CTG   ACG   ACT   CAG   CAT   CAT   CTG   GCC   CTG   ACG   ACT   CTG   ACG   ACT   CTG   ACG   ACT   CTG   CTG   CTG   CTG   CTG   ACG   ACT   ACG   ACG	CTG   GGT   TCT   TCA   GTT   CCT   GCC   CTG   ACG   ACG   CAA   CTG   CTG   CTG   ACG   CTG   ACG   CTG   ACG   CAT   CTG   CTG   CTG   ACG   ACG   CAT   CTG   CTG   CTG   ACG   ACG   CTG   ACG   CTG   ACG   CTG   CTG

FIGURE 9 1/5

390	TCA Ser>		GAT Asp>		GGC Gly>	TTG Leu>	280	GTC Val>	630	GTG Val>		GGA Gly>
	GCA Ala		AAT Asn		TTC Phe	530 CCT Pro	28	TTA		CTA		CGG Arg
•	TTA Leu	430	GCA Ala	480	CTT	AAT		GGT Gly		ATT Ile	0.29	GAT
380	AAT Asn	4	GAC		GAC	AAG Lys		TTG	620	AAT Asn	.9	ACC
•••	ACA Thr		GTA Val		GAG Glu	520 TGC AAA Cys Lys	570	GTG Val	Ū	AAC Asn		TGG Trp
	CTT Leu		CAG Gln	470	CCT	52 TGC Cys		TTT Phe		TTT Phe		GAC
370	AGT	420	GCA Ala	•	ACC Thr	GGC Gly	٠	GGA G1y	610	GGT Gly	¢ 660	GTT Val
, m	GAT Asp		ATG Met		TCT	CTT Leu	260	AGT	9	GGG		TAT Tyr
	AAA Lys		GAG Glu	460	ACT Thr	510 GCA Ala	u,	TGC		$_{\rm G1y}^{\rm GGT}$		CGG Arg
	GGT Gly	410	CTA Leu	4(	TGT	AAA Lys		GCA Ala		AGA	650	TCT Ser
360	TCA	7	GCT Ala		ATG Met	TCG	550	GCT Ala	* 009	ATT Ile	w	CTT
	CTC		AAA Lys		TTG	500 ATA Ile	5,	ACC Thr		CAC His		TCT Ser
	GTT Val	400	AGG Arg	450	GTT Val	CAG Gln		ATT Ile		TGC Cys	640	GAT Asp
350	AGG Arg	4(	GCA Ala		ATG Met	CCT		GAC Asp	290	GCT TGC Ala Cys	9	GCT
• •	CGA Arg		GCA Ala		GAT Asp	490 AGT GCT Ser Ala	540	TAC Tyr	<b>u</b> ,	GCT Ala		GGT Gly
	AAC Asn		GAG Glu	440	GTG Val	49 AGT Ser		TCT		TCA		ATT Ile

FIGURE 9
2/5

	TCA Ser>	GAT Asp>	0.3	GTT Val>	870	AGG Arg>		CGC Arg>		AAG Lys>	GCA Ala>
	CAG Gln	770 AGC Ser	82	GAA Glu		CCA		TTC Phe		GGA Gly	cAG CAG Gln
720	GTG Val	CAT His		GAT Asp		CCA	910	GTA Val	* 096	CTT	1010 CAT CAG His Gln
	GTG Val	TTG		GAA Glu	860	TTT Phe	91	GAG Glu		GCA	CTT
	GTA	760 TTT GAT Phe Asp	810	AAA Lys	ω	GAT Asp		AAA Lys		TCA	1000 TTG CTG Leu Leu
710	GCT	76 TTT Phe		ATC Ile		AGA Arg		$_{\rm G1y}^{\rm GGT}$	950	GAA Glu	1000 TTG C
•	GGA Gly	GCT		GCA Ala	850	TCC ATC Ser Ile	006	AAC Asn	O1	ATC Ile	TGG Trp
	GCT	TTT Phe	800	GCT Ala	8	TCC		ATG Met		TCA	GAC Asp
700	GAT GCT Asp Ala	750 CTC Leu	~	AAA Lys		$\frac{\text{GGG}}{\text{Gly}}$		CAA	940	CAG Gln	990 ATC Ile
7	GAT Asp	GGG G1y		CTA		AAT Asn	890	ATC Ile	6	CCT	AAC Asn
	GGA Gly	GAT Asp	790	CAT His	840	CAT His	w	TGC		GTG Val	TCC Ser
	TTT Phe	740 GAA Glu	7.	AGG Arg		GGA Gly		TCT Ser		TCT Ser	980 AAT GGA Asn Gly
069	CTC	GAG		CAA Gln		CTG	880	TAC	930	CGC Arg	9 AAT Asn
	ATT Ile	GCT		GGG G1y	830	GCC Ala	88	TCA		TGC	CTT Leu
	TGT Cys	730 ST GAT 'S ASP	780	GAT Asp	w	AAA Lys		TCT		GCT	'0 GGT Gly
680	ACA Thr	73 TGT Cys		GGA Gly		GAT Asp		CGT	920	TTT Phe	970 GCC GGT Ala Gly

FIGURE 9 3/5

1060	r ccr caa   Pro Gln>	1110	r GCG GCA : Ala Ala>		r GTG AAG n Val Lys>		ACA TGG	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT
1050	CGT CTA GAG GTT Arg Leu Glu Val	1100	AAC ACT AGT Asn Thr Ser	1150	AGG AGT GGA AAT Arg Ser Gly Asn	1200	GCC GGA CTC Ala Gly Leu	1250	GCCGAGCCAG	1310	CCANAAAAG AAGAAGTCAG	1370	CTTCATCACA TIGCCCTITI ICGIICCCCI
1040	GCA ACA CGT Ala Thr Arg	1090	AAT TAC GGG Asn Tyr Gly	1140	GCT GTG AGG Ala Val Arg	1190	GGA TTT GGC Gly Phe Gly	1240	ATT ATC AGG TGG GGA TAA GACTGAA GCCGAGCCAG CACTGCAGCT	1300	CCGATGTTTC ACGAAATTTT GCTTCCATGA	1360	CTTCATCACA
10	ATT GAT GCA GTA Ile Asp Ala Val	08*	TCA AAC TTG GCA Ser Asn Leu Ala	1130	CTA GAC GAA Leu Asp Glu	1180	GCA ACC GCA Ala Thr Ala	1230	AGG TGG GGA Arg Trp Gly	1290	ACGAAATTTT	1350	ACGACACGAT
1030	ATC Ile	1080	ATT ATC TCA AMILLE ILE SER AS	1120	CCC TTG GCA CT Pro Leu Ala Le	1170	GTG ATT Val Ile	1220	ATT ATC AC	1280	CCGATGTTTC	1340	AGCAAGCAAC ACGACACGAT
1020	AAT CAG AGG Asn Gln Arg	1070	GAA CGA ATI Glu Arg Ile	11	TCC ATT CCC Ser Ile Pro	1160	CCG GGT CAC Pro Gly His	1210	GGT TCT GCT Gly Ser Ala	1270	TCCTCTCAAA	1330	TCTTTTATGG

FIGURE 9

				1570 1580	1570
AAAAAAAAA	AAAAAAAA	TTTGCTAAAA	GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA AAAAAAAA	CATAAACATC	GAGATGACAG
1560	1550	1540	1530	1520	1510
CGGGACATTG	CATTTTGTCT	GCTTTTACTT	TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTACTT CATTTTGTCT CGGGACATTG	GTTTCTTGTT	TAAGTTATTT
1500	1490	1480	1470	1460	1450
TTGTCCCCAA	ATAGTTTCTT	TACAATACCC	TITCCATTAG ITTGATGATT TTGCTGACAA TACAATACCC ATAGTITCTT TTGTCCCCAA	TTTGATGATT	TTTCCATTAG
1440	1430	1420	1410	1400	1390

FIGURE 9 5/5

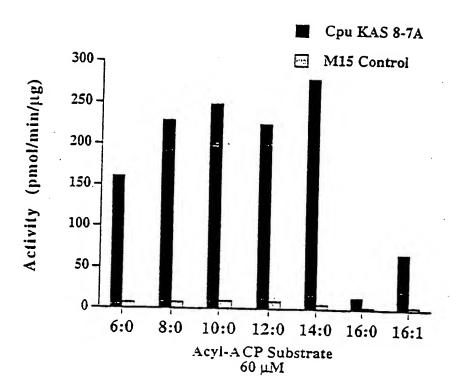


FIGURE 10

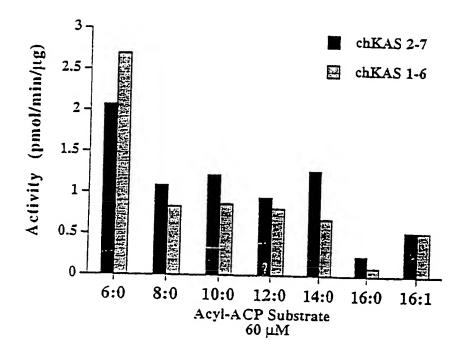


FIGURE 11

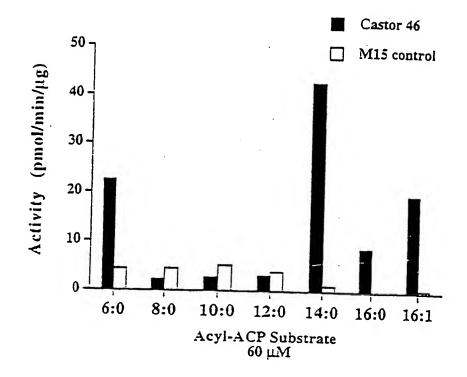
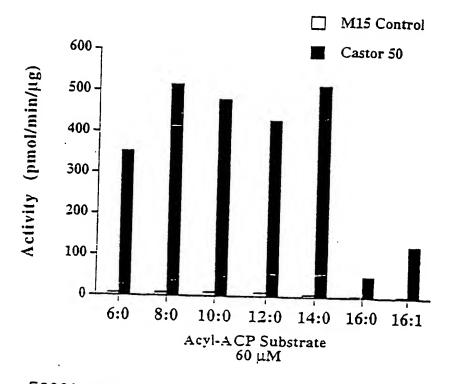
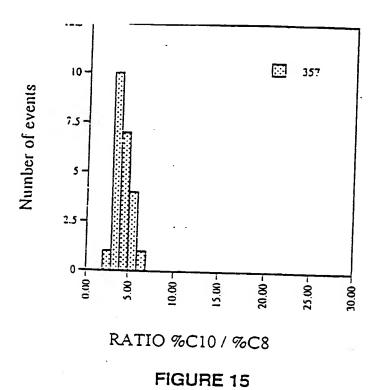


FIGURE 12



E328013-28

FIGURE 13



1/2

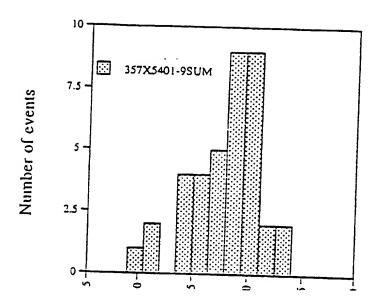
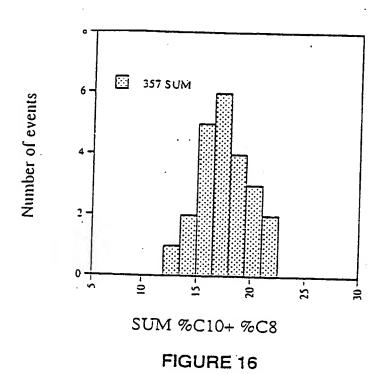


FIGURE 15 2/2



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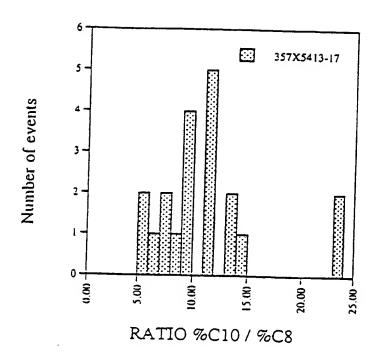


FIGURE 17 1/2

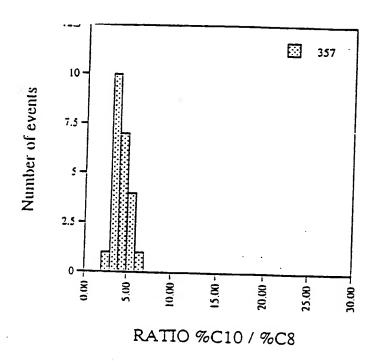


FIGURE 17

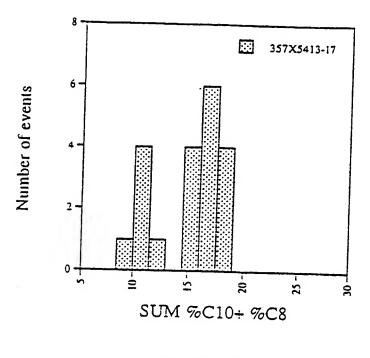
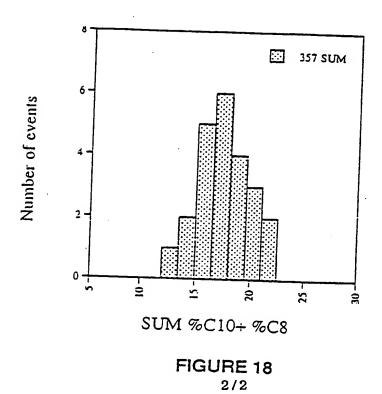
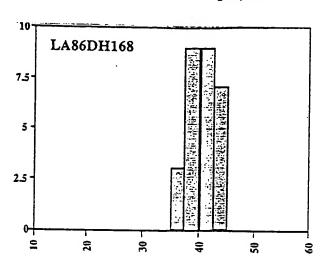


FIGURE 18 1/2



SUBSTITUTE SHEET (RULE 26)





## 12:0 levels (w%)

FIGURE 19 1/3

SUBSTITUTE SHEET (RULE 26)

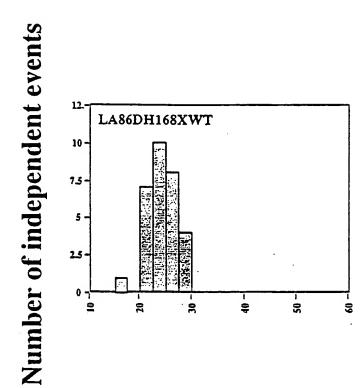
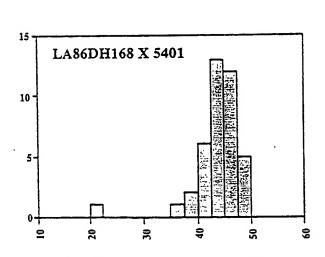


FIGURE 19 3/3 SUBSTITUTE SHEET (RULE 26)



12:0 levels (w%)

FIGURE 19 2/3

SUBSTITUTE SHEET (RULE 26)

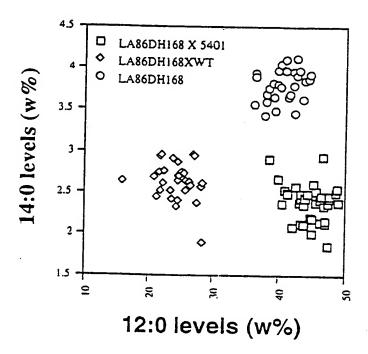
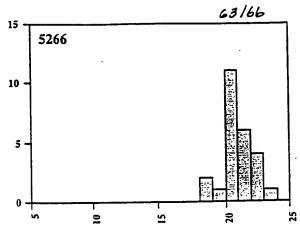


FIGURE 20

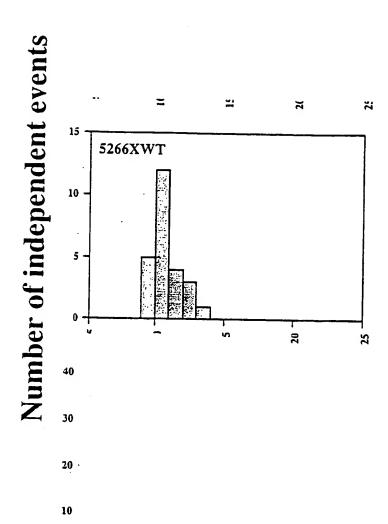
**WO 98/46776** 





## 18:0 levels (w%)

FIGURE -21. 1/3



18:0 levels (w%)

0

FIGURE 21. 2/3

SUBSTITUTE SHEET (RULE 26)

## Number of independent events

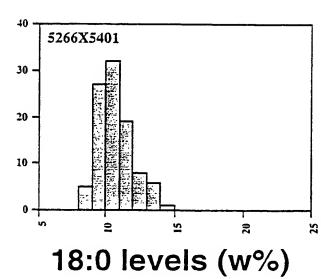


FIGURE 21

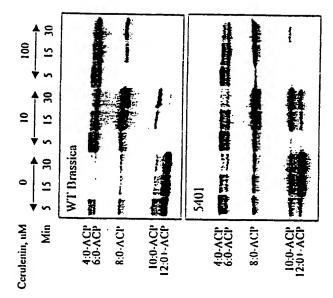


FIGURE 22

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